

<b>BTY 5001</b>	<b>PLANT TISSUE CULTURE TECHNIQUES</b> (Credit4;Theory 3hrs;Practical3 hrs)
<b>AIM</b>	To familiarize with plant tissue culture techniques
<b>Objectives</b>	<ul style="list-style-type: none"> <li>• To teach different components used in tissue culture media and their specific uses.</li> <li>• To teach different methods of micropropagation and their advantages</li> <li>• To teach different techniques used in in vitro conservation.</li> </ul>
<b>Learning outcome</b>	<p>After completion of the course, the students</p> <ul style="list-style-type: none"> <li>➤ Are able to understand the principles of plant tissue culture and various <i>in vitro</i> techniques</li> <li>➤ Proficient for developing haploid and triploid plants through tissue culture protocol.</li> <li>➤ Understand the techniques of protoplast isolation, culture and fusion and their application in crop improvements.</li> </ul>
<b>Sl.No</b>	<b>Theory</b>
1.	History of plant tissue culture, cellular totipotency: concepts and applications.
2.	Techniques of plant tissue culture, essential requirements of a plant tissue culture laboratory, Plant tissue culture media, General composition of the solid and liquid media, various gelling agents, media selection.
3.	Sterilization of medium, glasswares, instruments, plant material, transfer area, Preparation of explants, sterilization culture and incubation. Subculture and hardening. Micropropagation: various stages of micropropagation, importance.
4.	Principles and protocol applications of culture of different explants, embryo culture, importance of embryo culture
5.	Haploid plant production, Importance of haploid plants. Androgenesis: pre-treatment of anther/pollen grains, callus induction and shoot regeneration, androgenic embryos, their development. Merits and demerits of anther culture. Microspore culture, Protocol, Advantages of microspore culture over anther culture.
6.	<i>In vitro</i> gynogenesis, Ovary/ovule/flower bud culture, embryo induction from cultured ovary/ovule/flower bud, Callus induction from embryo sac cells and their organogenesis, advantages of gynogenetic plants over androgenic plants
7.	Triploid plant production: Importance of triploid plants, endosperm culture, stage of endosperm culture, role of embryo in endosperm culture, advantages and limitations of triploid plants.
8.	Suspension culture, batch culture, continuous culture, single cell culture.
9.	Somatic embryogenesis: Factors affecting somatic embryogenesis, differences between somatic and zygotic embryogenesis, synthetic seed production, desiccated and hydrated synthetic seeds, merits and demerits of synthetic seeds, somaclonal variation and applications of somaclonal variation in crop improvement.
10.	Protoplast isolation, culture, plant regeneration from protoplast, protoplast fusion and somatic hybridization, cybrids.
11.	In vitro germplasm storage, in-situ conservation, ex-situ conservation, cryopreservation.
12.	Application of tissue culture for crop improvement, problems, limitations and future prospectus.
<b>S. No.</b>	<b>Practical</b>
1.	Preparation of the stock solutions of MS medium,

2.	Preparation of MS medium from stock solutions,
3.	Isolation, preparation, sterilization and inoculation of different explants like shoot tip, node, anther, embryo and cambium
4.	Isolation and fusion of plant protoplasts,
5.	Preparation of synthetic seeds,
6.	Preparation of selective medium for drought or salinity resistance. Preparation of MS solid medium from stock solutions containing auxin and cytokinin, NaCl or PEG, and inoculation,
7.	Find out the uninucleate stage of anther and anther culture
8.	Dissect out an embryo from any seed and culture it on a suitable solid medium.

**Text Books:**

1. Barbara M. Reed (2008) Plant Cryopreservation: A Practical Guide. Springer, Heidelberg.
2. Bhojwani SS, Razdan MK (1996) Plant tissue culture: Theory and Practice. Elsevier, North Holland
3. Colin Ratledge, Bjorn Kristianson (2001) Basic biotechnology. Cambridge University press.
4. Dixon RA, Gonzales RA. (2004) Plant cell culture, a practical approach (II Edn). Oxford University Press.
5. Erica E. Benson (1999) Plant Conservation Biotechnology. Taylor and Francis, USA
6. Evans DE, Coleman JOD, Kearns A (2003) Plant Cell Culture. Taylor and Francis, USA.
7. Gamborg L, Philips GC (Eds.) (2005) Plant cell, tissue and organ culture: Fundamental methods. Narosa Publishing House, New Delhi.
8. Hamish A Collin, Sue Edwards (1998) Plant tissue culture. Bios scientific publishers, India
9. Michael R. Davey, Paul Anthony (2010) Plant Cell Culture: Essential Methods. Wiley-Blackwell Publishers, India
10. Susan R. Barnum (1998) Biotechnology an introduction. Wadsworth Publishing Company, USA.
11. Wang TL, Cuming A. (1996) Embryogenesis the generation of a plant. Bios Scientific Publishers Limited, UK
12. William J Thieman, Michael A Palladino (2009) Introduction to biotechnology (II Edn). Pearson.