

Mitochondrial DNA

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Muniyandi Nagarajan, Manikam Raja & Potnuru Vikram

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SHORT COMMUNICATION

Genetic characterization of *Bagarius* species using cytochrome *c* oxidase I and cytochrome *b* genesMuniyandi Nagarajan¹, Manikam Raja², and Potnuru Vikram¹¹Department of Genomic Science, School of Biological Sciences, Central University of Kerala, Kasaragod, Kerala, India and ²Department of Biotechnology, Periyar University, Salem, Tamil Nadu, India**Abstract**

In this study, we first inferred the genetic variability of two *Bagarius bagarius* populations collected from Ganges and Brahmaputra rivers of India using two mtDNA markers. Sequence analysis of *COI* gene did not show significant differences between two populations whereas cytochrome *b* gene showed significant differences between two populations. Followed by, genetic relationship of *B. bagarius* and *B. yarrielli* was analyzed using *COI* and cytochrome *b* gene and the results showed a higher level genetic variation between two species. The present study provides support for the suitability of *COI* and cytochrome *b* genes for the identification of *B. bagarius* and *B. yarrielli*.

Keywords

B. bagarius and *B. yarrielli*, *COI* gene, cytochrome *b* gene

History

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Introduction

Bagarius is one of the widely distributed fish genus in South Asian countries, which includes four species namely, *B. bagarius*, *B. rutilus*, *B. suchus*, and *B. yarrelli*. Among the four species, *B. bagarius* has got wide distribution in Indian river systems (Roberts, 1983). It was described from the Ganges, known locally by several names “Gorua”, “Baghar”, and “Bagari”. It is an important edible fish and fetches a high prize in the local markets due to its less spines and taste. The natural stocks of this species have declined drastically for the last two decades, which has seriously affected the livelihoods of local fishing communities (Allan et al., 2005). Subsequently, it has been categorized as a threatened species as per IUCN (Menon, 1989). Genetic variability analysis within and between populations can provide genetic relationship of the local populations, which is essential information for the effective fishery resource management. Among the molecular markers, cytochrome *c* oxidase I (*COI*) and cytochrome *b* genes are the widely used mitochondrial genes for studying intra and inter species genetic variability in fish (Habib et al., 2011; Luhariya et al., 2012; Mandal et al., 2012). In spite of its economic importance and conservation status, the genetic variability analysis on *Bagarius* species has not been conducted yet. In this study, the mtDNA *COI* and cytochrome *b* genes were used to examine the population structure of *B. bagarius* and genetic variability between *B. bagarius* and *B. yarrelli*.

Methods

A total of 26 *B. bagarius* samples were collected from Ganges river at Uttar Pradesh and Bihar (13 nos.) and Brahmaputra river

at Assam and Arunachal Pradesh (13 nos.). Genomic DNA was isolated from fin clips using Invitrogen’s “Pure Link Genomic DNA Mini Kit” following the manufactures instructions. The *COI* and cytochrome *b* genes were amplified with standard PCR conditions using the universal primers FishF1 and FishR1 (Ward et al., 2005) and L14724 and H15915 (Xiao et al., 2001), respectively. Sequencing was performed using Big Dye Terminator on ABI 3500 Genetic Analyzer (Applied Biosystems, Foster City, CA). In addition to that, the genetic variability between *B. bagarius* and *B. yarrielli* was analyzed using *COI* and cytochrome *b* gene sequences. The *COI* and cytochrome *b* gene sequences of *B. yarrielli* were retrieved from GenBank (Table 1). Sequences were edited and aligned using CLUSTAL X and MEGA software (Kumar et al., 2004). Haplotype diversity, AMOVA, and pair wise differences between populations (F_{ST}) were calculated using ARLEQUIN (Schneider et al., 2002). The phylogenetic network was drawn using the software NETWORK (Schneider et al., 2002).

Results**Genetic diversity between *B. bagarius* populations***COI* gene

The aligned 644 bp sequences of *COI* gene revealed seven unique haplotypes resulting from 10 polymorphic sites on 24 fish samples (Figure 1A). The Ganges and Brahmaputra populations showed four haplotypes each with seven and three polymorphic sites, respectively. The haplotype diversity was found to be 0.5256 ± 0.1527 and 0.4909 ± 0.1754 , respectively. Among the seven haplotypes, Hap3 was found in both the populations with high frequency (17 on 24) whereas others are unique to respective populations (Hap1, Hap2, and Hap4 observed in Ganges population; Hap5, Hap6, and Hap7 observed in Brahmaputra population). The AMOVA analysis showed 99.12% variation

within population whereas 0.88% was between populations. The pairwise F_{ST} indicated less variation (0.0088) between two populations.

Cytochrome *b* gene

Sequence data for the 913 bp regions of 25 fish resulted in four polymorphic sites, which defined seven unique haplotypes (Figure 1B). The Ganges population showed five haplotypes

Table 1. GenBank accession numbers for the mitochondrial COI and cytochrome *b* gene sequences used in this study.

S. no	Species	GenBank accession number	Gene
1.	<i>B. bagarius</i>	KP829955–KP829978	<i>COI</i>
		JN815268	
		JN697602	
		JN697601	
		EU417763	
2.	<i>B. bagarius</i>	EU417762	Cytochrome <i>b</i>
		FJ459434	
		KP829930–KP829954	
3.	<i>B. yarrelli</i>	HM156377–HM156380	<i>COI</i>
		JQ026260	
		EU490855	
4.	<i>B. yarrelli</i>	HQ322524	Cytochrome <i>b</i>
		HQ322525	
		AF416897	
		DQ119406	
		EU490904	
		JQ026260	

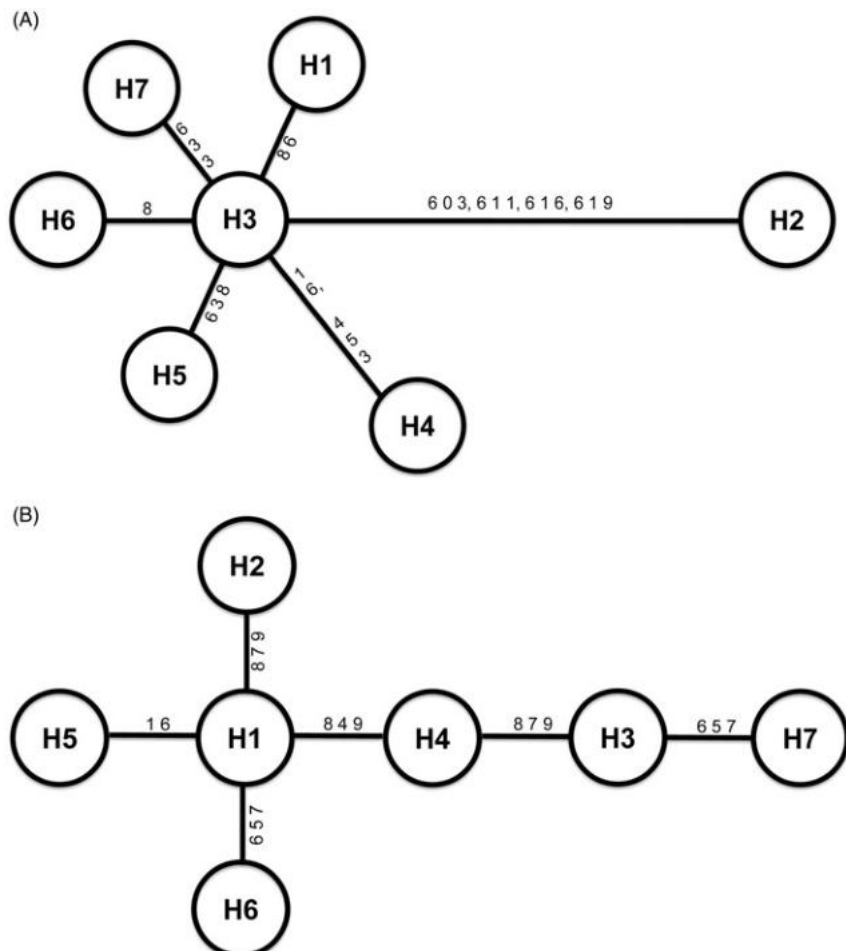
with three polymorphic sites where as the Brahmaputra population showed two haplotypes with two polymorphic sites. The haplotype diversity was found to be of 0.8030 ± 0.0903 and 0.1538 ± 0.1261 , respectively. Among the seven haplotypes, Hap1–Hap5 were found in Ganges population whereas Hap6 and Hap7 were found in Brahmaputra population. At nucleotide position 657, a “T” was prominent in Brahmaputra population instead “C”, which unambiguously separated both the populations. The AMOVA analysis showed 59.23% variation within population whereas 40.77% was between populations. In contrast to *COI* gene, the pairwise F_{ST} showed high differences (0.5923) between populations.

Genetic diversity between *B. bagarius* and *B. yarrelli*

COI gene

The sequences were truncated to 581 bp length for which sequences were available for *B. yarrelli*. Sequences data for the 581 bp regions of 36 fish revealed eight unique haplotypes resulting from 78 polymorphic sites. *B. bagarius* showed five haplotypes with five polymorphic sites whereas the *B. yarrelli* showed three haplotypes with 49 polymorphic sites. Among the eight haplotypes, Hap3 was found to be recurrent in *B. bagarius* (25 on 30). *B. yarrelli* with moderate sample size found to be divergent with high number of polymorphic sites and high haplotype diversity (0.6010 ± 0.2152) as compared with *B. bagarius* (0.3080 ± 0.1075). The AMOVA analysis showed 93.66% variation between species whereas 6.34% was within species. The pairwise F_{ST} values showed high level variation (0.9366) between *B. bagarius* and *B. yarrelli*.

Figure 1. Haplotype network for *COI* (A) and cytochrome *b* (B) genes of *B. bagarius*.



Cytochrome *b* gene

The 913 bp length cytochrome *b* gene revealed 12 unique haplotypes resulting from 132 polymorphic sites on 31 fishes. The *B. bagarius* showed seven haplotypes with four polymorphic sites whereas the *B. yarrelli* showed five haplotypes with 92 polymorphic sites. Among the seven haplotypes, Hap6 was found to be more frequent in *B. bagarius* (12 on 30). *Bagarius yarrelli* with moderate sample size was found to be divergent with a high number of polymorphic sites and high haplotype diversities (0.9333 ± 0.1217) as compared with *B. bagarius* (0.7637 ± 0.0774). The AMOVA analysis showed 85.08% variation between two species whereas within species was 14.92%. The pairwise F_{ST} values showed high level variation (0.8508) between *B. bagarius* and *B. yarrelli*.

Discussion

In this study, the genetic structure of *B. bagarius* was studied using two mitochondrial DNA markers. A total of 14 different haplotypes were found in *Bagarius* populations for COI and cytochrome *b* genes. However, only a single haplotype (Hap3 of the COI) was shared between two populations, which confirmed that these two populations were evolved from a common ancestor assuming Hap3 was the founding haplotype. The rest of the haplotypes would have evolved after the fragmentation. Among the two markers, cytochrome *b* gene showed high genetic variation between populations (59%) whereas COI gene showed high variation within population (99%). The genetic variation obtained in this study for within and between populations was higher than that of reported previously for migratory fishes (Vrijenhoek, 1998). The pairwise F_{ST} value was significantly high (0.59) between two *B. bagarius* populations for cytochrome *b* gene. Such high inter-population diversity can be expected as these two populations were collected from two different rivers (Habib et al., 2011; Ward et al., 1994). A nucleotide ‘‘T’’ was prominent in Brahmaputra population instead ‘‘C’’ at nucleotide position 657 in Cytochrome *b* gene, which unambiguously distinguish both the populations further which can be used as population specific marker in conservation studies. It indicates that currently there is no gene flow between Ganges and Brahmaputra river populations. The absence of gene flow further suggests that genetically isolated *B. bagarius* stocks exist in the Indian region. It has been reported that populations growing in different habitats are likely to diverge genetically from one another due to adaptive radiation (Greenwood, 1991). Mandal et al. (2009) have also observed similar results in *C. chitala* populations of Ganges and Brahmaputra. Followed by, COI and cytochrome *b* gene sequences have been used to characterize *B. bagarius* and *B. yarrelli*, which clearly distinguished both the species and strongly support the classification based on the morphological traits. The interspecific variation was greater than intraspecific variation, which is in agreement with the results reported previously for COI and cytochrome *b* genes (Habib et al., 2011; Nadiatul et al., 2011).

In conclusion, the present study clearly shows that the mtDNA COI and cytochrome *b* are found to be potential markers for studying the variations both among the population and the species. This study has provided basic information on the genetic

variability of *Bagarius* species, which can be used to develop strategies and management plan for the conservation of threatened *Bagarius* species.

Declaration of interest

The authors acknowledge the Science and Engineering Research Board (SERB), Department of Science and Technology, Government of India, New Delhi, for providing financial assistance (No. SR/FT/LS-40/2010). The authors declare that there are no competing interests. The authors alone are responsible for the content and writing of the paper.

References

- Allan JD, Abell R, Hogan Z, Revenga C, Taylor BW, Welcome RI, Winemiller K. (2005). Over fishing of inland waters. *Bioscience* 55: 1041–50.
- Greenwood PH. (1991). Speciation in cichlid fishes. In: Keenleyside MHA, editor. *Cichlid fishes behaviour, ecology and evolution*. London: Chapman & Hall. p 86–102.
- Habib M, Lakra WS, Mohindra V, Khare P, Barman AS, Singh A, Lal KK, et al. (2011). Evaluation of cytochrome *b* mtDNA sequences in genetic diversity studies of *Channa marulius* (Channidae: Perciformes). *Mol Biol Rep* 3:841–6.
- Kumar S, Tamura K, Nei M. (2004). MEGA 3.1: Integrated software for molecular evolutionary genetics analysis and sequence alignment. *Brief Bioinformatics* 5:150–63.
- Luhariya RK, Lal KK, Singh RK, Mohindra V, Punia P, Chauhan UK, Gupta A, Lakra WS. (2012). Genetic divergence in wild population of *Labeo rohita* (Hamilton, 1822) from nine Indian rivers, analyzed through MtDNA cytochrome *b* region. *Mol Biol Rep* 39:3659–65.
- Mandal A, Lal KK, Mohindra V, Singh RK, Punia P, Chauhan UK, Lakra WS. (2009). Evaluation of genetic variation in the Clown Knifefish, *Chitala chitala*, using allozymes, RAPD, and microsatellites. *Biochem Genet* 47:216–34.
- Mandal A, Mohindra V, Singh RK, Punia P, Singh AK, Lal KK. (2012). Mitochondrial DNA variation in natural populations of endangered Indian feather-back fish, *Chitala chitala*. *Mol Biol Rep* 39:1765–75.
- Menon AGK. (1989). Conservation of Ichthyofauna of India. In: Jhingran AG, Sugunan VV, editors. *Conservation and management of inland capture fisheries resources of India*. India: The Inland Fisheries Society of India.
- Nadiatul HH, Daud SK, Siraj SS, Sungan S, Moghaddam FY. (2011). Genetic diversity of Malaysian indigenous Mahseer, *Tor douronensis* in Sarawak river basins as revealed by cytochrome *c* oxidase I gene sequences. *Iran J Anim Biosyst* 7:119–27.
- Ng H, Kottelat M. (2000). Descriptions of three new species of catfishes (Teleostei: Akysidae and Sisoridae) from Laos and Vietnam. *J South Asian Nat Hist* 5:7–15.
- Roberts T. (1983). Revision of the South and Southeast Asian Sisorid Catfish Genus *Bagarius*, with description of a new species from the Mekong. *Copeia* 2:435–45.
- Schneider S, Roessli D, Excoffier L. (2002). ARLEQUIN 2.0: A software for population genetic data analysis. Switzerland: Genetics and Biometry Laboratory, University of Geneva.
- Vrijenhoek RC. (1998). Conservation genetics of freshwater fish. *J Fish Biol* 53:394–412.
- Xiao WY, Zhang H, Liu (2001). Molecular systematics of *Xenocyprinae* (Teleostei: Cyprinidae): Taxonomy, biogeography, and coevolution of a special group restricted in East Asia. *Mol Phylogenet Evol* 18:163–73.
- Ward RD, Woodward M, Skibinski DOF. (1994). A comparison of genetic diversity levels in marine, freshwater, and anadromous fishes. *J Fish Biol* 44:213–32.
- Ward RD, Zemlak TS, Innes BH, Last PR, Hebert PDN. (2005). DNA barcoding Australia’s fish species. *Phil Trans R Soc B* 360:1847–57.